A1847-LUC ΔHER2 cells were generated from the human ovarian cancer cell line A1847 by CRISPR followed by clonal selection and transduction with replication-defective lentivirus encoding luciferase. The loss of surface HER2 expression was confirmed by flow cytometry (Figure 1) and expression of luciferase was confirmed by luminescence after exposure to luciferin (Figure 2).

Storage

Store vial in liquid nitrogen immediately upon receipt.

Formulation

Cells are cryopreserved in 0.5 ml - 1 ml of 90% FBS + 10% DMSO.

Thaw Protocol

Partially immerse the vial in a 37°C water bath with gentle shaking until most of the medium is thawed. In a tissue culture hood, add 1 ml of pre-warmed culture medium into the vial and immediately transfer the contents of the vial to a centrifuge tube containing 5-10 ml of pre-warmed culture medium. Centrifuge the tube at room temperature for 5 minutes, aspirate the supernatant and suspend the cell pellet in 5-10 ml of pre-warmed culture medium.

Culture Protocol

A1847-LUC ΔEpCAM is an adherent cell line. Culture the cells in RPMI-1640 medium containing 10% FBS using a humidified incubator set to 5% CO_2. When the cell monolayer is nearly confluent, use trypsin-EDTA to detach the cells from the culture flask and immediately neutralize the trypsin by adding fresh, pre-warmed culture medium to the cells. Cells should be passaged at least twice per week.

Notices & Disclaimer

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Loss of HER2 Expression

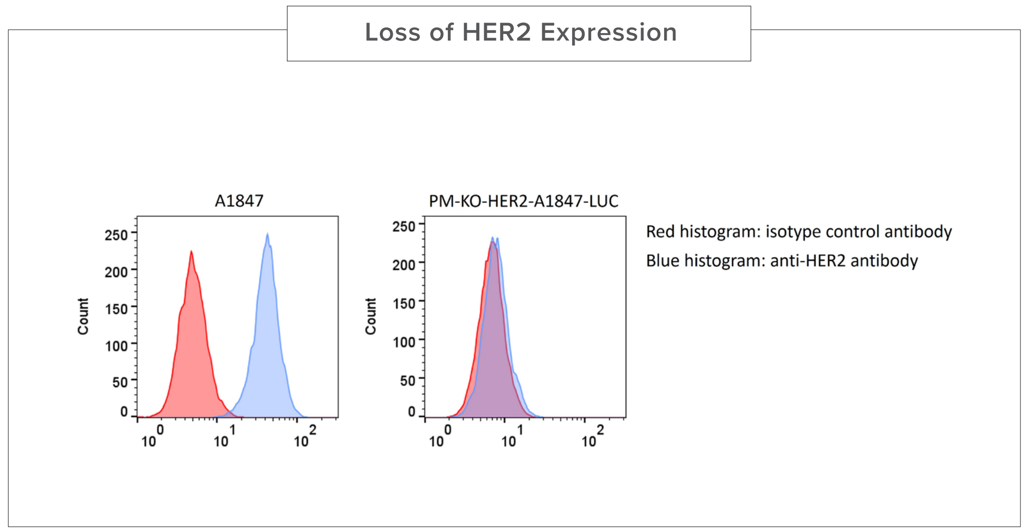

Figure 1. PM-KO-HER2-A1847-LUC cells and parental A1847 cells were stained with an antibody specific for HER2 and an isotype control antibody. The HER2 antibody bound to A1847 cells but did not bind to PM-KO-HER2-A1847-LUC cells.

Expression of luciferase

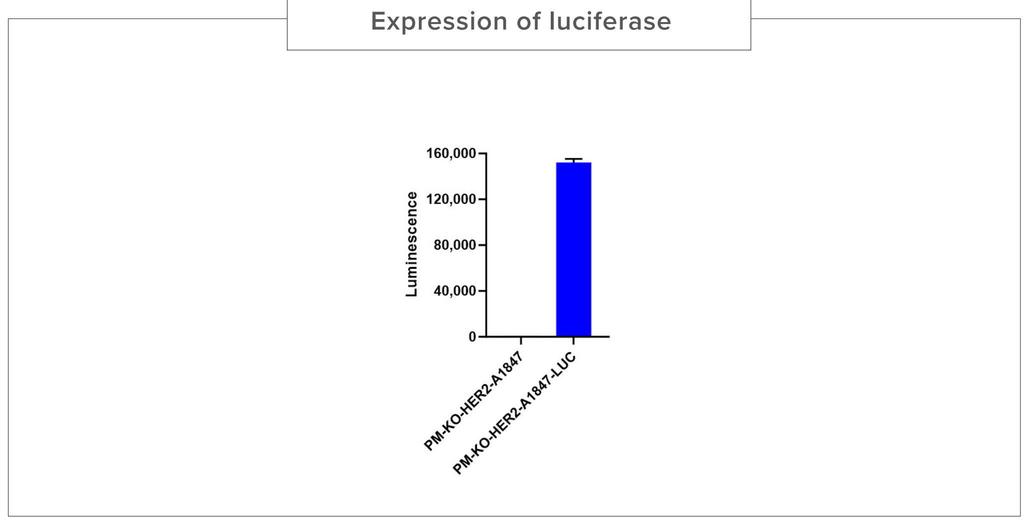

Figure 2. PM-KO-HER2-A1847-LUC cells and control PM-KO-HER2-A1847 cells were lysed and incubated with luciferin to assess luciferase expression. Only PM-KO-HER2-A1847-LUC cells became luminescent.

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