

CT26-EpCAM cells were generated from the BALB/c mouse colon cancer cell line CT26 by transduction with replication-defective lentivirus encoding human EpCAM. Surface expression of EpCAM was confirmed by flow cytometry (Figure 1).

<b>Storage</b>
Store vial in liquid nitrogen immediately upon receipt.
<b>Formulation</b>
Cells are cryopreserved in 0.5 ml - 1 ml of 90% FBS + 10% DMSO.
<b>Thaw Protocol</b>
Partially immerse the vial in a 37°C water bath with gentle shaking until most of the medium is thawed. In a tissue culture hood, add 1 ml of pre-warmed culture medium into the vial and immediately transfer the contents of the vial to a centrifuge tube containing 5-10 ml of pre-warmed culture medium. Centrifuge the tube at room temperature for 5 minutes, aspirate the supernatant and suspend the cell pellet in 5-10 ml of pre-warmed culture medium.
<b>Culture Protocol</b>
CT26-EpCAM is an adherent cell line. Culture the cells in RPMI-1640 medium containing 10% FBS using a humidified incubator set to 5% CO <sub>2</sub> . When the cell monolayer is nearly confluent, use trypsin-EDTA to detach the cells from the culture flask and immediately neutralize the trypsin by adding fresh, pre-warmed culture medium to the cells. Cells should be passaged at least twice per week.
<b>Notices &amp; Disclaimer</b>
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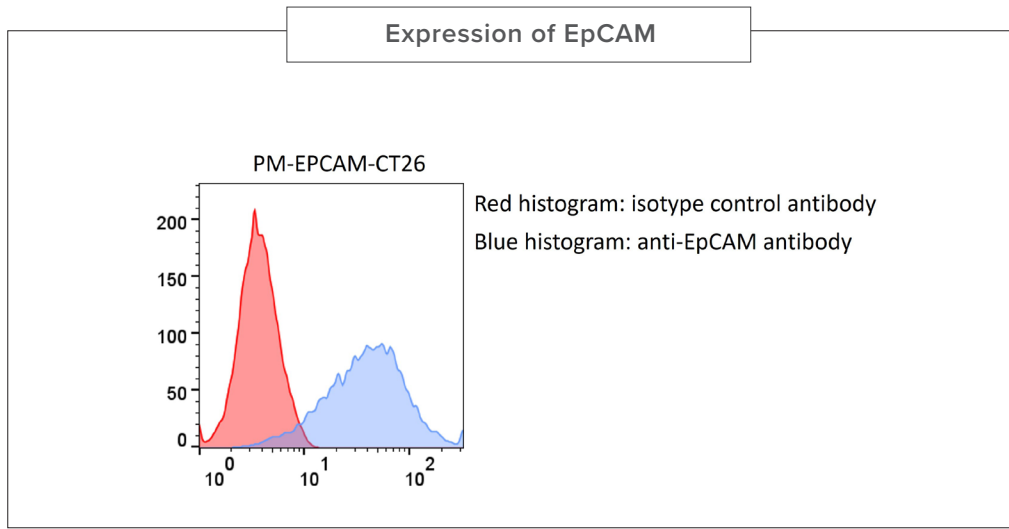


Figure 1. PM-EPCAM-CT26 cells were stained with an antibody specific for EpCAM and an isotype control antibody. The EpCAM antibody bound to the PM-EPCAM-CT26 cells.

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