HeLa-CD19 cells were generated from the human cervical cancer cell line HeLa by transduction with replication-defective lentivirus encoding human CD19. Surface expression of CD19 was confirmed by flow cytometry (Figure 1).

Storage

Store vial in liquid nitrogen immediately upon receipt.

Formulation

Cells are cryopreserved in 0.5 ml - 1 ml of 90% FBS + 10% DMSO.

Thaw Protocol

Partially immerse the vial in a 37°C water bath with gentle shaking until most of the medium is thawed. In a tissue culture hood, add 1 ml of pre-warmed culture medium into the vial and immediately transfer the contents of the vial to a centrifuge tube containing 5-10 ml of pre-warmed culture medium. Centrifuge the tube at room temperature for 5 minutes, aspirate the supernatant and suspend the cell pellet in 5-10 ml of pre-warmed culture medium.

Culture Protocol

HeLa-CD19 is an adherent cell line. Culture the cells in DMEM containing 10% FBS using a humidified incubator set to 5% CO2. When the cell monolayer is nearly confluent, use trypsin-EDTA to detach the cells from the culture flask and immediately neutralize the trypsin by adding fresh, pre-warmed culture medium to the cells. Cells should be passaged at least twice per week.

Notices & Disclaimer

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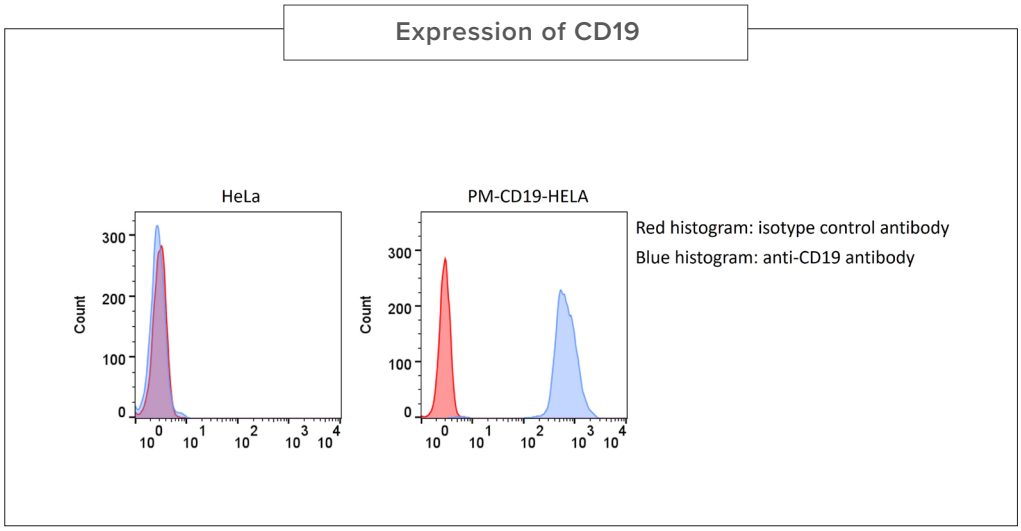

Figure 1. PM-CD19-HELA cells and parental HeLa cells were stained with an antibody specific for CD19 and an isotype control antibody. The CD19 antibody bound to PM-CD19-HELA cells but not to HeLa cells.

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